

# GLAD

April 20, 2011

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arrayCGH	<i>Object of Class arrayCGH</i>
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## Description

Description of the object arrayCGH.

## Value

The object arrayCGH is a list with at least a data.frame named arrayValues and a vector named arrayDesign. The data.frame arrayValues must contain the following fields :

Col	Vector of columns coordinates.
Row	Vector of rows coordinates.
...	Other elements can be added.

The vector arrayDesign is composed of 4 values : c(arrayCol, arrayRow, SpotCol, SpotRow). The array CGH is represented by arrayRow\*arrayCol blocs and each bloc is composed of SpotRow\*SpotCol spots.

N.B. : Col takes the values in 1:arrayRow\*SpotRow and Row takes the values in 1:arrayCol\*SpotCol

## Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

## Author(s)

Philippe Hupé, <glad@curie.fr>.

## See Also

[glad](#).

**Examples**

```
data(arrayCGH)

# object of class arrayCGH

array <- list(arrayValues=array2, arrayDesign=c(4,4,21,22))
class(array) <- "arrayCGH"
```

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arrayPersp

*Perspective image of microarray spots statistic*


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**Description**

The function `arrayPersp` creates perspective images of shades of gray or colors that correspond to the values of a statistic for each spot on the array. The statistic can be the intensity log-ratio, a spot quality measure (e.g. spot size or shape), or a test statistic. This function can be used to explore whether there are any spatial effects in the data, for example, print-tip or cover-slip effects.

**Usage**

```
## Default S3 method:
arrayPersp(Statistic, Col, Row,
           ArrCol, ArrRow, SpotCol, SpotRow,
           mediancenter=FALSE,
           col=myPalette("green", "red", "yellow"),
           zlim=zlim, bar=TRUE, ...)

## S3 method for class 'arrayCGH':
arrayPersp(arrayCGH, variable,
           mediancenter=FALSE,
           col=myPalette("green", "red", "yellow"),
           zlim=zlim, bar=TRUE, ...)
```

**Arguments**

<code>arrayCGH</code>	Object of class <code>arrayCGH</code> .
<code>variable</code>	Variable to be plotted
<code>Statistic</code>	Statistic to be plotted.
<code>Col</code>	Vector of columns coordinates.
<code>Row</code>	Vector of rows coordinates.
<code>ArrCol</code>	Number of columns for the blocs.
<code>ArrRow</code>	Number of rows for the blocs.
<code>SpotCol</code>	Number of column for each bloc.
<code>SpotRow</code>	Number of rows for each bloc.

mediancenter	If mediancenter=TRUE, values of <code>Statistic</code> are median-centered.
col	List of colors such as that generated by <code>Palettes</code> . In addition to these color palettes functions, a new function <code>myPalette</code> was defined to generate color palettes from user supplied low, middle, and high color values.
zlim	Numerical vector of length 2 giving the extreme values of <code>z</code> to associate with colors <code>low</code> and <code>high</code> of <code>myPalette</code> . By default <code>zlim</code> is the range of <code>z</code> . Any values of <code>z</code> outside the interval <code>zlim</code> will be truncated to the relevant limit.
bar	If <code>bar=TRUE</code> , a calibration color bar is shown to the right of the image.
...	Graphical parameters can be given as arguments to function <code>persp</code> . N.B. : <code>Col</code> takes the values in <code>1:arrayRow*SpotRow</code> and <code>Row</code> takes the values in <code>1:arrayCol*SpotCol</code>

### Value

An image is created on the current graphics device.

### Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

### Author(s)

Philippe Hupé, <glad@curie.fr>.

### See Also

`persp`, `arrayPlot`, `myPalette`.

### Examples

```
data(arrayCGH)

# object of class arrayCGH

array <- list(arrayValues=array2, arrayDesign=c(4,4,21,22))
class(array) <- "arrayCGH"

arrayPersp(array, "Log2Rat", main="Perspective image of array CGH",
            box=FALSE, theta=110, phi=40)
```

arrayPlot

*Spatial image of microarray spots statistic***Description**

The function `arrayPlot` creates spatial images of shades of gray or colors that correspond to the values of a statistic for each spot on the array. The statistic can be the intensity log-ratio, a spot quality measure (e.g. spot size or shape), or a test statistic. This function can be used to explore whether there are any spatial effects in the data, for example, print-tip or cover-slip effects.

**Usage**

```
## Default S3 method:
arrayPlot(Statistic, Col, Row,
          ArrCol, ArrRow, SpotCol, SpotRow,
          mediancenter=FALSE,
          col=myPalette("green", "red", "yellow"),
          contour=FALSE, nlevels=5,
          zlim=NULL, bar=c("none", "horizontal", "vertical"),
          layout=TRUE, ...)

## S3 method for class 'arrayCGH':
arrayPlot(arrayCGH, variable,
          mediancenter=FALSE,
          col=myPalette("green", "red", "yellow"),
          contour=FALSE, nlevels=5,
          zlim=NULL, bar=c("none", "horizontal", "vertical"),
          layout=TRUE, ...)
```

**Arguments**

<code>arrayCGH</code>	Object of class <code>arrayCGH</code> .
<code>variable</code>	Variable to be plotted
<code>Statistic</code>	Statistic to be plotted.
<code>Col</code>	Vector of columns coordinates.
<code>Row</code>	Vector of rows coordinates.
<code>ArrCol</code>	Number of columns for the blocs.
<code>ArrRow</code>	Number of rows for the blocs.
<code>SpotCol</code>	Number of column for each bloc.
<code>SpotRow</code>	Number of rows for each bloc.
<code>mediancenter</code>	If <code>mediancenter=TRUE</code> , values of <code>Statistic</code> are median-centered.
<code>col</code>	List of colors such as that generated by <code>Palettes</code> . In addition to these color palettes functions, a new function <code>myPalette</code> was defined to generate color palettes from user supplied low, middle, and high color values.
<code>contour</code>	If <code>contour=TRUE</code> , contour are plotted, otherwise they are not shown.
<code>nlevels</code>	Numbers of levels added by <code>contour</code> if <code>contour=TRUE</code> .

<code>zlim</code>	Numerical vector of length 2 giving the extreme values of <code>z</code> to associate with colors <code>low</code> and <code>high</code> of <code>myPalette</code> . By default <code>zlim</code> is the range of <code>z</code> . Any values of <code>z</code> outside the interval <code>zlim</code> will be truncated to the relevant limit.
<code>bar</code>	If <code>bar=='horizontal'</code> (resp. <code>'vertical'</code> ), an horizontal (resp. vertical) calibration color bar is shown to the right of the image.
<code>layout</code>	If <code>layout==TRUE</code> plot layout is automatically set when a color bar is asked for
<code>...</code>	Graphical parameters can be given as arguments to function <code>image</code> . N.B. : <code>Col</code> takes the values in <code>1:arrayRow*SpotRow</code> and <code>Row</code> takes the values in <code>1:arrayCol*SpotCol</code>

### Details

This function is very similar to the `maImage` written by Sandrine Dudoit (available in `marrayPlots` package) with added options `zlim`, `mediancenter` and `layout`.

### Value

An image is created on the current graphics device.

### Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

### Author(s)

Philippe Hupé, <glad@curie.fr>.

### See Also

`image`, `contour`, `arrayPersp`, `myPalette`.

### Examples

```
data(arrayCGH)

pdf(file="arrayCGH.pdf", height=21/cm(1), width=29.7/cm(1))
arrayPlot(array2$Log2Rat, array2$Col, array2$Row, 4,4,21,22, main="Spatial Image of array
dev.off()

# object of class arrayCGH

array <- list(arrayValues=array2, arrayDesign=c(4,4,21,22))
class(array) <- "arrayCGH"

arrayPlot(array, "Log2Rat", main="Spatial Image of array CGH")
```

---

```
as.data.frame.profileCGH
      profileCGH consercion
```

---

### Description

Convert a profileCGH object into a data.frame.

### Usage

```
## S3 method for class 'profileCGH':
as.data.frame(x, row.names = NULL, optional = FALSE, ...)
```

### Arguments

x	The object to converted into data.frame.
row.names	NULL or a character vector giving the row names for the data frame. Missing values are not allowed.
optional	logical. If 'TRUE', setting row names and converting column names (to syntactic names) is optional.
...	...

### Details

The attributes `profileValues` and `profileValuesNA` are binded into a data.frame.

### Value

A data.frame object

### Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

### Author(s)

Philippe Hupé, <glad@curie.fr>

### See Also

[as.profileCGH](#)

### Examples

```
data(snijders)

### Creation of "profileCGH" object
profileCGH <- as.profileCGH(gm13330)
```

```
#####
###
### glad function as described in Hupé et al. (2004)
###
#####

res <- glad(profileCGH, mediancenter=FALSE,
            smoothfunc="lawsglad", bandwidth=10, round=2,
            model="Gaussian", lkern="Exponential", qlambda=0.999,
            base=FALSE,
            lambdabreak=8, lambdacluster=8, lambdaclusterGen=40,
            type="tricubic", param=c(d=6),
            alpha=0.001, msize=5,
            method="centroid", nmax=8,
            verbose=FALSE)

res <- as.data.frame(res)
```

---

as.profileCGH

*Create an object of class profileCGH*


---

## Description

Create an object of class profileCGH.

## Usage

```
as.profileCGH(object, ...)
## S3 method for class 'data.frame':
as.profileCGH(object, infaction=c("value","empty"), value=20, ...)
```

## Arguments

object	A data.frame to be convert into profileCGH.
infaction	If "value" then the LogRatio with infinite values (-Inf, Inf) are replace by + or - value according to the sign. If "empty" then NAs are put instead.
value	replace Inf by value if infaction is "value".
...	...

## Details

The data.frame to be convert must at least contain the following fields: LogRatio, PosOrder, and Chromosome. If the field Chromosome is of mode character, it is automatically converted into a numeric vector (see [ChrNumeric](#)); a field ChromosomeChar contains the character labels. The data.frame to be converted into a profileCGH objet is split into two data.frame: profileValuesNA contains the rows for which there is at least a missing value for either LogRatio, PosOrder or Chromosome; profileValues contains the remaining rows.

**Value**

A list with the following attributes

```
profileValues      A data.frame
profileValuesNA    A data.frame
```

**Note**

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

**Author(s)**

Philippe Hupé, <glad@curie.fr>

**See Also**

[as.data.frame.profileCGH](#)

**Examples**

```
data(snijders)

### Creation of "profileCGH" object
profileCGH <- as.profileCGH(gm13330)

attributes(profileCGH)
```

---

array

*Bladder cancer CGH data*

---

**Description**

Bladder cancer data from 3 arrays CGH (Comparative Genomic Hybridization). Arrays dimension are 4 blocs per column, 4 blocs per row, 21 columns per bloc and 22 rows by blocs.

**Usage**

```
data(arrayCGH)
```

**Format**

A data frame composed of the following elements :

**Log2Rat** Log 2 ratio.

**Position** BAC position on the genome.

**CHROMOSOME** Chromosome.

**Col** Column location on the array.

**Row** Row location on the array.



**Source**

Institut Curie, <glad@curie.fr>.

**Examples**

```
data(arrayCGH)
data <- array1 #array1 to array3
```

---

ChrNumeric

*Convert chromosome into numeric values*

---

**Description**

Convert chromosome into numeric values.

**Usage**

```
ChrNumeric(Chromosome)
```

**Arguments**

Chromosome    A vector with chromosome labels.

**Details**

For sexual chromosome, labels must contains X or Y which are coded by 23 and 24 respectively.

**Note**

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

**Author(s)**

Philippe Hupé, <glad@curie.fr>

**Examples**

```
Chromosome <- c("1", "X", "Y", "chr X", "ChrX", "chrX", "Chr    Y")
ChrNumeric(Chromosome)
```

ColorBar

*Calibration bar for color images***Description**

This function produces a color image (color bar) which can be used for the legend to another color image obtained from the functions `image` or `arrayPlot`.

**Usage**

```
ColorBar(x, horizontal=TRUE, col=heat.colors(50), scale=1:length(x), k=10, ...)
```

**Arguments**

<code>x</code>	If "numeric", a vector containing the "z" values in the color image, i.e., the values which are represented in the color image. Otherwise, a "character" vector representing colors.
<code>horizontal</code>	If TRUE, the values of <code>x</code> are represented as vertical color strips in the image, else, the values are represented as horizontal color strips.
<code>col</code>	Vector of colors such as that generated by <code>rainbow</code> , <code>heat.colors</code> , <code>topo.colors</code> , <code>terrain.colors</code> , or similar functions. In addition to these color palette functions, a new function <code>myPalette</code> was defined to generate color palettes from user supplied low, middle, and high color values.
<code>scale</code>	A "numeric" vector specifying the "z" values in the color image. This is used when the argument <code>x</code> is a "character" vector representing color information.
<code>k</code>	Object of class "numeric", for the number of labels displayed on the bar.
<code>...</code>	Optional graphical parameters, see <code>par</code> .

**Author(s)**

Sandrine Dudoit, Yee Hwa (Jean) Yang.

**See Also**

`image`, `arrayPlot` `myPalette`.

**Examples**

```
par(mfrow=c(3,1))
Rcol <- myPalette(low="white", high="red", k=10)
Gcol <- myPalette(low="white", high="green", k=50)
RGcol <- myPalette(low="green", high="red", k=100)
ColorBar(Rcol)
ColorBar(Gcol, scale=c(-5,5))
ColorBar(1:50, col=RGcol)

par(mfrow=c(1,3))
x<-seq(-1, 1, by=0.01)
ColorBar(x, col=Gcol, horizontal=FALSE, k=11)
ColorBar(x, col=Gcol, horizontal=FALSE, k=21)
```

```
ColorBar(x, col=Gcol, horizontal=FALSE, k=51)
```

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cytoband	<i>Cytogenetic banding</i>
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### Description

Cytogenetic banding

### Usage

```
data(cytoband)
```

### Examples

```
data(cytoband)
cytoband
```

---

daglad	<i>Analysis of array CGH data</i>
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### Description

This function allows the detection of breakpoints in genomic profiles obtained by array CGH technology and affects a status (gain, normal or lost) to each clone.

### Usage

```
## S3 method for class 'profileCGH':
daglad(profileCGH, mediancenter=FALSE,
normalrefcenter=FALSE, genomestep=FALSE,
OnlySmoothing = FALSE, OnlyOptimCall = FALSE,
smoothfunc="lawsglad", lkern="Exponential",
model="Gaussian", qlambda=0.999, bandwidth=10,
sigma=NULL, base=FALSE, round=2,
lambdabreak=8, lambdaclusterGen=40, param=c(d=6),
alpha=0.001, msize=5, method="centroid", nmin=1, nmax=8, region.size=2,
amplicon=1, deletion=-5, deltaN=0.10, forceGL=c(-0.15,0.15),
nbsigma=3, MinBkpWeight=0.35, DelBkpInAmp=TRUE,
CheckBkpPos=TRUE, assignGNLOut=TRUE,
breaksFdrQ = 0.0001, haarStartLevel = 1,
haarEndLevel = 5, weights.name = NULL,
verbose=FALSE, ...)
```

**Arguments**

<code>profileCGH</code>	Object of class <code>profileCGH</code>
<code>mediancenter</code>	If TRUE, <code>LogRatio</code> are center on their median.
<code>genomestep</code>	If TRUE, a smoothing step over the whole genome is performed and a "clustering throughout the genome" allows to identify a cluster corresponding to the Normal DNA level. The threshold used in the <code>daglad</code> function ( <code>deltaN</code> , <code>forceGL</code> , <code>amplicon</code> , <code>deletion</code> ) and then compared to the median of this cluster.
<code>normalrefcenter</code>	If TRUE, the <code>LogRatio</code> are centered through the median of the cluster identified during the <code>genomestep</code> .
<code>OnlySmoothing</code>	If TRUE, only segmentation is performed without optimization of breakpoints and calling.
<code>OnlyOptimCall</code>	If TRUE, the user can provide data which have been already segmented. In this case, <code>profileCGH\$profileValues</code> must contain a field with the name "Smoothing". The <code>daglad</code> function skip the smoothing step but both the optimization of breakpoints and calling are performed.
<code>smoothfunc</code>	Type of algorithm used to smooth <code>LogRatio</code> by a piecewise constant function. Choose either <code>lawsglad</code> , <code>haarseg</code> , <code>aws</code> or <code>laws</code> ( <code>aws</code> package).
<code>lkern</code>	<code>lkern</code> determines the location kernel to be used (see <code>laws</code> in <code>aws</code> package for details).
<code>model</code>	<code>model</code> determines the distribution type of <code>LogRatio</code> (see <code>laws</code> in <code>aws</code> package for details).
<code>qlambda</code>	<code>qlambda</code> determines the scale parameter <code>qlambda</code> for the stochastic penalty (see <code>laws</code> in <code>aws</code> package for details).
<code>base</code>	If TRUE, the position of clone is the physical position onto the chromosome, otherwise the rank position is used.
<code>sigma</code>	Value to be passed to either argument <code>sigma2</code> of <code>aws</code> (see <code>aws</code> package) function or <code>shape</code> of <code>laws</code> (see <code>aws</code> package). If NULL, <code>sigma</code> is calculated from the data.
<code>bandwidth</code>	Set the maximal bandwidth <code>hmax</code> in the <code>aws</code> or <code>laws</code> functions in <code>aws</code> package. For example, if <code>bandwidth=10</code> then the <code>hmax</code> value is set to $10 * X_N$ where $X_N$ is the position of the last clone.
<code>round</code>	The smoothing results of either <code>aws</code> or <code>laws</code> functions (in <code>aws</code> package) are rounded or not depending on the <code>round</code> argument. The <code>round</code> value is passed to the argument <code>digits</code> of the <code>round</code> function.
<code>lambdabreak</code>	Penalty term ( $\lambda'$ ) used during the "Optimization of the number of breakpoints" step.
<code>lambdaclusterGen</code>	Penalty term ( $\lambda^*$ ) used during the "clustering throughout the genome" step.
<code>param</code>	Parameter of kernel used in the penalty term.
<code>alpha</code>	Risk <code>alpha</code> used for the "Outlier detection" step.
<code>msize</code>	The outliers MAD are calculated on regions with a cardinality greater or equal to <code>msize</code> .

method	The agglomeration method to be used during the "clustering throughout the genome" steps.
nmin	Minimum number of clusters ( $N \cdot \max$ ) allowed during the "clustering throughout the genome" clustering step.
nmax	Maximum number of clusters ( $N \cdot \max$ ) allowed during the "clustering throughout the genome" clustering step.
region.size	The breakpoints which define regions with a number of probes lower or equal to this value are discarded.
amplicon	Level (and outliers) with a smoothing value (log-ratio value) greater than this threshold are consider as amplicon. Note that first, the data are centered on the normal reference value computed during the "clustering throughout the genome" step.
deletion	Level (and outliers) with a smoothing value (log-ratio value) lower than this threshold are consider as deletion. Note that first, the data are centered on the normal reference value computed during the "clustering throughout the genome" step.
deltaN	Region with smoothing values in between the interval $[-\delta N, +\delta N]$ are supposed to be normal.
forceGL	Level with smoothing value greater (lower) than <code>rangeGL[1]</code> ( <code>rangeGL[2]</code> ) are considered as gain (lost). Note that first, the data are centered on the normal reference value computed during the "clustering throughout the genome" step.
nbsigma	For each breakpoints, a weight is calculated which is a function of absolute value of the Gap between the smoothing values of the two consecutive regions. $Weight = 1 - \text{kernelpen}(\text{abs}(\text{Gap}), \text{param} = c(d = \text{nbsigma} \cdot \text{Sigma}))$ where Sigma is the standard deviation of the LogRatio.
MinBkpWeight	Breakpoints which <code>GNLchange==0</code> and Weight less than <code>MinBkpWeight</code> are discarded.
DelBkpInAmp	If TRUE, the breakpoints identified inside amplicon regions are deleted. For amplicon, the log-ratio values are highly variable which lead to identification of false positive breakpoints.
CheckBkpPos	If TRUE, the accuracy position of each breakpoints is checked.
assignGNLOut	If FALSE the status (gain/normal/loss) is not assigned for outliers.
breaksFdrQ	<code>breaksFdrQ</code> for HaarSeg algorithm.
haarStartLevel	<code>haarStartLevel</code> for HaarSeg algorithm.
haarEndLevel	<code>haarEndLevel</code> for HaarSeg algorithm.
weights.name	The name of the fields which contains the weights used for the haarseg algorithm. By default, the value is set to NULL meaning that all the observations have the same weights. If provided, the field must contain positive values.
verbose	If TRUE some information are printed.
...	...

## Details

The function `daglad` implements a slightly modified version of the methodology described in the article : Analysis of array CGH data: from signal ratio to gain and loss of DNA regions (Hupé et al., Bioinformatics, 2004). For smoothing, it is possible to use either the AWS algorithm (Polzehl

and Spokoiny, 2002) or the HaarSeg algorithm (Ben-Yaacov and Eldar, Bioinformatics, 2008). The daglad function allows to choose some threshold to help the algorithm to identify the status of the genomic regions. The thresholds are given in the following parameters:

- deltaN
- forceGL
- deletion
- amplicon

### Value

An object of class "profileCGH" with the following attributes:

profileValues

is a data.frame with the following information:

- **Smoothing**The smoothing values correspond to the median of each Level
- **Breakpoints**The last position of a region with identical amount of DNA is flagged by 1 otherwise it is 0. Note that during the "Optimization of the number of breakpoints" step, removed breakpoints are flagged by -1.
- **Level**Each position with equal smoothing value are labelled the same way with an integer value starting from one. The label is incremented by one when a new level occurs or when moving to the next chromosome.
- **OutliersAws**Each AWS outliers are flagged -1 (if it is in the  $\alpha/2$  lower tail of the distribution) or 1 (if it is in the  $\alpha/2$  upper tail of the distribution) otherwise it is 0.
- **OutliersMad**Each MAD outliers are flagged -1 (if it is in the  $\alpha/2$  lower tail of the distribution) or 1 (if it is in the  $\alpha/2$  upper tail of the distribution) otherwise it is 0.
- **OutliersTot**OutliersAws + OutliersMad.
- **NormalRef**Clusters which have been used to set the normal reference during the "clustering throughout the genome" step are code by 0. Note that if genomestep=FALSE, all the value are set to 0.
- **ZoneGNL**Status of each clone: Gain is coded by 1, Loss by -1, Amplicon by 2, deletion by -10 and Normal by 0.

BkpInfo

is a data.frame sum up the information for each breakpoint:

- **Chromosome**Chromosome name.
- **Smoothing**Smoothing value for the breakpoint.
- **Gap**absolute value of the gap between the smoothing values of the two consecutive regions.
- **Sigma**The estimation of the standard-deviation of the chromosome.
- **Weight1** - kernelpen(Gap, type, param=c(d=nbsigma\*Sigma))
- **ZoneGNL**Status of the level where is the breakpoint.
- **GNLchange**Takes the value 1 if the ZoneGNL of the two consecutive regions are different.
- **LogRatio**Test over Reference log-ratio.

NormalRef

If genomestep=TRUE and normalrefcenter=FALSE, then NormalRef is the median of the cluster which has been used to set the normal reference during the "clustering throughout the genome" step. Otherwise NormalRef is 0.

**Note**

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

**Author(s)**

Philippe Hupé, <glad@curie.fr>.

**References**

Hupé et al. (Bioinformatics, 2004): Analysis of array CGH data: from signal ratio to gain and loss of DNA regions.

Polzehl and Spokoiny (WIAS-Preprint 787, 2002): Local likelihood modelling by adaptive weights smoothing.

Ben-Yaacov and Eldar (Bioinformatics, 2008): A fast and flexible method for the segmentation of aCGH data.

**See Also**

[glad](#).

**Examples**

```
data(snijders)
gm13330$Clone <- gm13330$BAC
profileCGH <- as.profileCGH(gm13330)

#####
###
### daglad function
###
#####

res <- daglad(profileCGH, mediancenter=FALSE, normalrefcenter=FALSE, genomestep=FALSE,
              smoothfunc="lawsglad", lkern="Exponential", model="Gaussian",
              qlambda=0.999, bandwidth=10, base=FALSE, round=1.5,
              lambdabreak=8, lambdaclusterGen=40, param=c(d=6), alpha=0.001, msize=5,
              method="centroid", nmin=1, nmax=8,
              amplicon=1, deletion=-5, deltaN=0.10, forceGL=c(-0.15,0.15), nbsigma=3,
              MinBkpWeight=0.35, CheckBkpPos=TRUE)

### data for cytoband
data(cytoband)

### Genomic profile on the whole genome
plotProfile(res, unit=3, Bkp=TRUE, labels=FALSE, Smoothing="Smoothing",
            main="Breakpoints detection: DAGLAD analysis", cytoband = cytoband)

###Genomic profile for chromosome 1
```

```

plotProfile(res, unit=3, Bkp=TRUE, labels=TRUE, Chromosome=1,
Smoothing="Smoothing", main="Chromosome 1: DAGLAD analysis", cytoband = cytoband)

### The standard-deviation of LogRatio are:
res$SigmaC

### The list of breakpoints is:
res$BkpInfo

```

glad

*Analysis of array CGH data***Description**

This function allows the detection of breakpoints in genomic profiles obtained by array CGH technology and affects a status (gain, normal or lost) to each clone.

**Usage**

```

## S3 method for class 'profileCGH':
glad(profileCGH, mediancenter=FALSE,
      smoothfunc="lawsglad", bandwidth=10, round=1.5,
      model="Gaussian", lkern="Exponential", qlambda=0.999,
      base=FALSE, sigma,
      lambdabreak=8, lambdacluster=8, lambdaclusterGen=40,
      type="tricubic", param=c(d=6),
      alpha=0.001, msize=5,
      method="centroid", nmax=8, assignGNLOut=TRUE,
      breaksFdrQ = 0.0001, haarStartLevel = 1, haarEndLevel = 5,
      verbose=FALSE, ...)

```

**Arguments**

profileCGH	Object of class <code>profileCGH</code>
mediancenter	If TRUE, LogRatio are centered on their median.
smoothfunc	Type of algorithm used to smooth LogRatio by a piecewise constant function. Choose either <code>lawsglad</code> , <code>haarseg</code> , <code>aws</code> or <code>laws</code> in <code>aws</code> package.
bandwidth	Set the maximal bandwidth <code>hmax</code> in the <code>aws</code> or <code>laws</code> functions in <code>aws</code> package. For example, if <code>bandwidth=10</code> then the <code>hmax</code> value is set to $10 * X_N$ where $X_N$ is the position of the last clone.
round	The smoothing results are rounded or not depending on the <code>round</code> argument. The <code>round</code> value is passed to the argument <code>digits</code> of the <code>round</code> function.
model	Determines the distribution type of the LogRatio. Keep always the model as "Gaussian" (see <code>laws</code> in <code>aws</code> package).
lkern	Determines the location kernel to be used (see <code>aws</code> or <code>laws</code> in <code>aws</code> package).



qlambda	Determines the scale parameter for the stochastic penalty (see <code>aws</code> or <code>laws</code> in <code>aws</code> package)
base	If <code>TRUE</code> , the position of clone is the physical position on the chromosome, otherwise the rank position is used.
sigma	Value to be passed to either argument <code>sigma2</code> of <code>aws</code> function or shape of <code>laws</code> (see <code>aws</code> package). If <code>NULL</code> , <code>sigma</code> is calculated from the data.
lambdabreak	Penalty term ( $\lambda'$ ) used during the <b>Optimization of the number of breakpoints</b> step.
lambdacluster	Penalty term ( $\lambda^*$ ) used during the <b>MSHR clustering by chromosome</b> step.
lambdaclusterGen	Penalty term ( $\lambda^*$ ) used during the <b>HCSR clustering throughout the genome</b> step.
type	Type of kernel function used in the penalty term during the <b>Optimization of the number of breakpoints</b> step, the <b>MSHR clustering by chromosome</b> step and the <b>HCSR clustering throughout the genome</b> step.
param	Parameter of kernel used in the penalty term.
alpha	Risk alpha used for the <b>Outlier detection</b> step.
msize	The outliers MAD are calculated on regions with a cardinality greater or equal to <code>msize</code> .
method	The agglomeration method to be used during the <b>MSHR clustering by chromosome</b> and the <b>HCSR clustering throughout the genome</b> clustering steps.
nmax	Maximum number of clusters ( $N^*max$ ) allowed during the the <b>MSHR clustering by chromosome</b> and the <b>HCSR clustering throughout the genome</b> clustering steps.
assignGNLOut	If <code>FALSE</code> the status (gain/normal/loss) is not assigned for outliers.
breaksFdrQ	<code>breaksFdrQ</code> for HaarSeg algorithm.
haarStartLevel	<code>haarStartLevel</code> for HaarSeg algorithm.
haarEndLevel	for HaarSeg algorithm.
verbose	If <code>TRUE</code> some information are printed
...	...

## Details

The function `glad` implements the methodology which is described in the article: Analysis of array CGH data: from signal ratio to gain and loss of DNA regions (Hupé et al., Bioinformatics, 2004).

The principles of the GLAD algorithm: First, the detection of breakpoints is based on the estimation of a piecewise constant function with the Adaptive Weights Smoothing (AWS) procedure (Polzehl and Spokoiny, 2002). Alternatively, it is possible to use the HaarSeg algorithm (Ben-Yaacov and Eldar, Bioinformatics, 2008). Then, a procedure based on penalized maximum likelihood optimizes the number of breakpoints and removes the undesirable breakpoints. Finally, based on the regions previously identified, a two-step unsupervised classification (**MSHR clustering by chromosome** and the **HCSR clustering throughout the genome**) with model selection criteria allows a status to be assigned for each region (gain, loss or normal).

Main parameters to be tuned:

`qlambda` if you want the smoothing to fit some very local effect, choose a smaller `qlambda`.

bandwidth	choose a bandwidth not too small otherwise you will have a lot of little discontinuities.
lambdabreak	The higher the parameter is, the higher the number of undesirable breakpoints is.
lambdacluster	The higher the parameter is, the higher is the number of the regions within a chromosome which
lambdaclusterGen	More the parameter is high more the regions over the whole genome are supposed to belong to

## Value

An object of class "profileCGH" with the following attributes:

profileValues:

a data.frame with the following added information:

- **Smoothing**The smoothing values correspond to the median of each **MSHR (i.e. Region)**.
- **Breakpoints**The last position of a region with identical amount of DNA is flagged by 1 otherwise it is 0. Note that during the "Optimization of the number of breakpoints" step, removed breakpoints are flagged by -1.
- **Region**Each position between two breakpoints are labelled the same way with an integer value starting from one. The label is incremented by one when a new breakpoint is found or when moving to the next chromosome. The variable `region` is what we call MSHR.
- **Level**Each position with equal smoothing value is labelled the same way with an integer value starting from one. The label is incremented by one when a new level is found or when moving to the next chromosome.
- **OutliersAws**Each AWS outliers are flagged -1 or 1 otherwise it is 0.
- **OutliersMad**Each MAD outliers are flagged -1 (if it is in the  $\alpha/2$  lower tail of the distribution) or 1 (if it is in the  $\alpha/2$  upper tail of the distribution) otherwise it is 0.
- **OutliersTot**OutliersAws + OutliersMad.
- **ZoneChr**Clusters identified after **MSHR (i.e. Region) clustering by chromosome**.
- **ZoneGen**Clusters identified after **HCSR clustering throughout the genome**.
- **ZoneGNL**Status of each clone : Gain is coded by 1, Loss by -1 and Normal by 0.

BkpInfo:

the data.frame attribute `BkpInfo` which gives the list of breakpoints:

- **PosOrder**The rank position of each clone on the genome.
- **PosBase**The base position of each clone on the genome.
- **Chromosome**Chromosome name.

SigmaC:

the data.frame attribute `SigmaC` gives the estimation of the LogRatio standard-deviation for each chromosome:

- **Chromosome**Chromosome name.
- **Value**The estimation is based on the Inter Quartile Range.

## Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

## Author(s)

Philippe Hupé, <glad@curie.fr>.

## References

- Hupé et al. (Bioinformatics, 2004) Analysis of array CGH data: from signal ratio to gain and loss of DNA regions.
- Polzehl and Spokoiny (WIAS-Preprint 787, 2002) Local likelihood modelling by adaptive weights smoothing.
- Ben-Yaacov and Eldar (Bioinformatics, 2008) A fast and flexible method for the segmentation of aCGH data.

## See Also

[profileCGH](#), [as.profileCGH](#), [plotProfile](#).

## Examples

```
data(snijders)

### Creation of "profileCGH" object
gm13330$Clone <- gm13330$BAC
profileCGH <- as.profileCGH(gm13330)

#####
###
### glad function as described in Hupé et al. (2004)
###
#####

res <- glad(profileCGH, mediancenter=FALSE,
            smoothfunc="lawsglad", bandwidth=10, round=1.5,
            model="Gaussian", lkern="Exponential", qlambda=0.999,
            base=FALSE,
            lambdabreak=8, lambdacluster=8, lambdaclusterGen=40,
            type="tricubic", param=c(d=6),
            alpha=0.001, msize=5,
            method="centroid", nmax=8,
            verbose=FALSE)

### cytoband data to plot chromosomes
data(cytoband)

### Genomic profile on the whole genome
plotProfile(res, unit=3, Bkp=TRUE, labels=FALSE, Smoothing="Smoothing",
main="Breakpoints detection: GLAD analysis", cytoband = cytoband)

### Genomic profile for chromosome 1
plotProfile(res, unit=3, Bkp=TRUE, labels=TRUE, Chromosome=1,
Smoothing="Smoothing", main="Chromosome 1: GLAD analysis", cytoband = cytoband)

### The standard-deviation of LogRatio are:
res$SigmaC

### The list of breakpoints is:
```

```
res$BkpInfo
```

---

```
hclustglad
```

*Hierarchical Clustering*

---

## Description

Hierarchical cluster analysis on a set of dissimilarities and methods for analyzing it.

## Usage

```
hclustglad(d, method = "complete", members=NULL)
```

## Arguments

<code>d</code>	a dissimilarity structure as produced by <code>dist</code> .
<code>method</code>	the agglomeration method to be used. This should be (an unambiguous abbreviation of) one of "ward", "single", "complete", "average", "mcquitty", "median" or "centroid".
<code>members</code>	NULL or a vector with length size of <code>d</code> .

## Details

This function performs a hierarchical cluster analysis using a set of dissimilarities for the  $n$  objects being clustered. Initially, each object is assigned to its own cluster and then the algorithm proceeds iteratively, at each stage joining the two most similar clusters, continuing until there is just a single cluster. At each stage distances between clusters are recomputed by the Lance–Williams dissimilarity update formula according to the particular clustering method being used.

A number of different clustering methods are provided. *Ward's* minimum variance method aims at finding compact, spherical clusters. The *complete linkage* method finds similar clusters. The *single linkage* method (which is closely related to the minimal spanning tree) adopts a ‘friends of friends’ clustering strategy. The other methods can be regarded as aiming for clusters with characteristics somewhere between the single and complete link methods.

If `members != NULL`, then `d` is taken to be a dissimilarity matrix between clusters instead of dissimilarities between singletons and `members` gives the number of observations per cluster. This way the hierarchical cluster algorithm can be “started in the middle of the dendrogram”, e.g., in order to reconstruct the part of the tree above a cut (see examples). Dissimilarities between clusters can be efficiently computed (i.e., without `hclustglad` itself) only for a limited number of distance/linkage combinations, the simplest one being squared Euclidean distance and centroid linkage. In this case the dissimilarities between the clusters are the squared Euclidean distances between cluster means.

In hierarchical cluster displays, a decision is needed at each merge to specify which subtree should go on the left and which on the right. Since, for  $n$  observations there are  $n - 1$  merges, there are  $2^{(n-1)}$  possible orderings for the leaves in a cluster tree, or dendrogram. The algorithm used in `hclustglad` is to order the subtree so that the tighter cluster is on the left (the last, i.e. most recent, merge of the left subtree is at a lower value than the last merge of the right subtree). Single observations are the tightest clusters possible, and merges involving two observations place them in order by their observation sequence number.

**Value**

An object of class **hclust** which describes the tree produced by the clustering process. The object is a list with components:

merge	an $n - 1$ by 2 matrix. Row $i$ of <code>merge</code> describes the merging of clusters at step $i$ of the clustering. If an element $j$ in the row is negative, then observation $-j$ was merged at this stage. If $j$ is positive then the merge was with the cluster formed at the (earlier) stage $j$ of the algorithm. Thus negative entries in <code>merge</code> indicate agglomerations of singletons, and positive entries indicate agglomerations of non-singletons.
height	a set of $n - 1$ non-decreasing real values. The clustering <i>height</i> : that is, the value of the criterion associated with the clustering <code>method</code> for the particular agglomeration.
order	a vector giving the permutation of the original observations suitable for plotting, in the sense that a cluster plot using this ordering and matrix <code>merge</code> will not have crossings of the branches.
labels	labels for each of the objects being clustered.
call	the call which produced the result.
method	the cluster method that has been used.
dist.method	the distance that has been used to create <code>d</code> (only returned if the distance object has a "method" attribute).

**Author(s)**

The `hclustglad` function is based an Algorithm contributed to STATLIB by F. Murtagh.

**References**

- Everitt, B. (1974). *Cluster Analysis*. London: Heinemann Educ. Books.
- Hartigan, J. A. (1975). *Clustering Algorithms*. New York: Wiley.
- Sneath, P. H. A. and R. R. Sokal (1973). *Numerical Taxonomy*. San Francisco: Freeman.
- Anderberg, M. R. (1973). *Cluster Analysis for Applications*. Academic Press: New York.
- Gordon, A. D. (1999). *Classification*. Second Edition. London: Chapman and Hall / CRC
- Murtagh, F. (1985). "Multidimensional Clustering Algorithms", in *COMPSTAT Lectures 4*. Wuerzburg: Physica-Verlag (for algorithmic details of algorithms used).

**Examples**

```
data(USArrests)
hc <- hclustglad(dist(USArrests), "ave")
plot(hc)
plot(hc, hang = -1)

## Do the same with centroid clustering and squared Euclidean distance,
## cut the tree into ten clusters and reconstruct the upper part of the
## tree from the cluster centers.
hc <- hclustglad(dist(USArrests)^2, "cen")
memb <- cutree(hc, k = 10)
cent <- NULL
for(k in 1:10){
```

```
cent <- rbind(cent, colMeans(USArrests[memb == k, , drop = FALSE]))
}
hc1 <- hclustglad(dist(cent)^2, method = "cen", members = table(memb))
opar <- par(mfrow = c(1, 2))
plot(hc, labels = FALSE, hang = -1, main = "Original Tree")
plot(hc1, labels = FALSE, hang = -1, main = "Re-start from 10 clusters")
par(opar)
```

---

kernelpen

*Kernelpen function*

---

### Description

Kernel function used in the penalty term.

### Usage

```
kernelpen(x, type="tricubic", param)
```

### Arguments

x	Real Value.
type	Type of kernelpen to be used
param	a named vector.

### Details

The only kernel available is the "tricubic" kernel which takes the values  $(1 - (x/d)^3)^3$ . The value of d is given by `param=c(d=6)` for example.

### Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

### Author(s)

Philippe Hupé, <glad@curie.fr>

---

`myPalette`*Microarray color palette*

---

### Description

This function returns a vector of color names corresponding to a range of colors specified in the arguments.

### Usage

```
myPalette(low = "white", high = c("green", "red"), mid=NULL, k =50)
```

### Arguments

<code>low</code>	Color for the lower end of the color palette, specified using any of the three kinds of R colors, i.e., either a color name (an element of <code>colors</code> ), a hexadecimal string of the form <code>"#rrggbb"</code> , or an integer <code>i</code> meaning <code>palette()[i]</code> .
<code>high</code>	Color for the upper end of the color palette, specified using any of the three kinds of R colors, i.e., either a color name (an element of <code>colors</code> ), a hexadecimal string of the form <code>"#rrggbb"</code> , or an integer <code>i</code> meaning <code>palette()[i]</code> .
<code>mid</code>	Color for the middle portion of the color palette, specified using any of the three kinds of R colors, i.e., either a color name (an element of <code>colors</code> ), a hexadecimal string of the form <code>"#rrggbb"</code> , or an integer <code>i</code> meaning <code>palette()[i]</code> .
<code>k</code>	Number of colors in the palette.

### Value

A "character" vector of color names. This can be used to create a user-defined color palette for subsequent graphics by `palette`, in a `col=` specification in graphics functions, or in [par](#).

### Author(s)

Sandrine Dudoit, Yee Hwa (Jean) Yang.

### See Also

`palette`, `rgb`, `colors`, `col2rgb`, [image](#), [ColorBar](#), [arrayPlot](#).

### Examples

```
par(mfrow=c(1,4))
pal <- myPalette(low="red", high="green")
ColorBar(seq(-2,2, 0.2), col=pal, horizontal=FALSE, k=21)
pal <- myPalette(low="red", high="green", mid="yellow")
ColorBar(seq(-2,2, 0.2), col=pal, horizontal=FALSE, k=21)
pal <- myPalette()
ColorBar(seq(-2,2, 0.2), col=pal, horizontal=FALSE, k=21)
pal <- myPalette(low="purple", high="purple", mid="white")
ColorBar(seq(-2,2, 0.2), col=pal, horizontal=FALSE, k=21)
```

---

plotProfile *Plot genomic profile and cytogenetic banding*

---

### Description

Plot genomic profile with breakpoints, outliers, smoothing line and cytogenetic banding.

### Usage

```
## S3 method for class 'profileCGH':
plotProfile(profileCGH, variable="LogRatio", Chromosome=NULL,
            Smoothing=NULL, GNL="ZoneGNL", Bkp=FALSE,
            labels=TRUE, plotband=TRUE, unit=0,
            coldAGLAD=c("black", "blue", "red", "green", "yellow"),
            pchSymbol=c(20, 13),
            colCytoBand=c("white", "darkblue"),
            colCentro="red", text=NULL, cytoband = NULL, main="", ylim)
```

### Arguments

profileCGH	Object of class <code>profileCGH</code>
variable	The variable to be plot.
Chromosome	A numeric vector with chromosome number to be plotted. Use 23 and 24 for chromosome X and Y respectively. If <code>NULL</code> , all the genome is plotted.
Smoothing	The variable used to plot the smoothing line. If <code>NULL</code> , nothing is plotted.
GNL	The variable used to plot the Gain, Normal and Loss color code.
Bkp	If <code>TRUE</code> , the breakpoints are represented by a vertical red dashed line.
labels	If <code>TRUE</code> , the labels of the cytogenetic banding are written.
plotband	If <code>TRUE</code> , the cytogenetic banding are plotted.
unit	Give the unit of the PosBase. For example if <code>unit=3</code> , PosBase are in Kb, if <code>unit=6</code> , PosBase are in Mb, ...
coldAGLAD	Color code to plot Deletion, Amplification, Gain, Lost and Normal status.
pchSymbol	A vector of two elements to specify the symbol tu be used for plotting point. <code>pchSymbol[2]</code> is the symbol for outliers.
colCytoBand	Color code for cytogenetic banding.
colCentro	Color code for centromere.
text	A list with the parameters to be passed to the function <code>text</code> .
cytoband	cytodand data. For human, cytoband data are avaibale using <code>data(cytoband)</code> .
main	title of the plot.
ylim	range of the y-axis
...	...



**Details**

" "

**Value**

A plot

**Note**

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

**Author(s)**

Philippe Hupé, &lt;glad@curie.fr&gt;.

**See Also**

" "

**Examples**

```
### Cytogenetic banding information
data(cytoband)

###
data(snijders)

### Creation of "profileCGH" object
profileCGH <- as.profileCGH(gm13330)

#####
###
### glad function as described in Hupé et al. (2004)
###
#####

res <- glad(profileCGH, mediancenter=FALSE,
            smoothfunc="lawsglad", bandwidth=10, round=2,
            model="Gaussian", lkern="Exponential", qlambda=0.999,
            base=FALSE,
            lambdabreak=8, lambdacluster=8, lambdaclusterGen=40,
            type="tricubic", param=c(d=6),
            alpha=0.001, msize=5,
            method="centroid", nmax=8,
            verbose=FALSE)

### cytoband data to plot chromosome
data(cytoband)

### Genomic profile on the whole genome
plotProfile(res, unit=3, Bkp=TRUE, labels=FALSE,
```

```

Smoothing="Smoothing", plotband=FALSE, cytoband = cytoband)

### Genomic profile on the whole genome and cytogenetic banding
plotProfile(res, unit=3, Bkp=TRUE, labels=FALSE, Smoothing="Smoothing",
            cytoband = cytoband)

### Genomic profile for chromosome 1
text <- list(x=c(90000,200000),y=c(0.15,0.3),labels=c("NORMAL","GAIN"), cex=2)
plotProfile(res, unit=3, Bkp=TRUE, labels=TRUE, Chromosome=1,
            Smoothing="Smoothing", plotband=FALSE, text=text, cytoband = cytoband)

### Genomic profile for chromosome 1 and cytogenetic banding with labels
text <- list(x=c(90000,200000),y=c(0.15,0.3),labels=c("NORMAL","GAIN"), cex=2)
plotProfile(res, unit=3, Bkp=TRUE, labels=TRUE, Chromosome=1,
            Smoothing="Smoothing", text=text, main="Chromosome 1", cytoband = cytoband)

```

---

profileCGH

*Objects of Class profileCGH and profileChr*


---

## Description

Description of the objects `profileCGH` and `profileChr`. The last object corresponds to data of only one chromosome.

## Details

`LogRatio`, `Chromosome` and `PosOrder` are compulsory.

## Value

Objects `profileCGH` and `profileChr` are composed of a list with the first element `profileValues` which is a `data.frame` with the following columns names:

<code>LogRatio</code>	Test over Reference log-ratio.
<code>PosOrder</code>	The rank position of each clone on the genome.
<code>PosBase</code>	The base position of each clone on the genome.
<code>Chromosome</code>	Chromosome name.
<code>Clone</code>	The name of the corresponding clone.
<code>...</code>	Other elements can be added.

## Note

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

## Author(s)

Philippe Hupé, <glad@curie.fr>.

**See Also**

`glad`, `as.profileCGH`.

**Examples**

```
data(snijders)
gm13330$Clone <- gm13330$BAC
profileCGH <- as.profileCGH(gm13330)
class(profileCGH) <- "profileCGH"

profileChr <- as.profileCGH(gm13330[which(gm13330$Chromosome==1),])
class(profileChr) <- "profileChr"
```

---

snijders

*Public CGH data of Snijders*

---

**Description**

The data consist of 15 human cell strains with known karyotype (12 fibroblast cell strains, 2 chorionic villus cell strains, 1 lymphoblast cell strain) from the NIGMS Human Genetics Cell Repository (<http://locus.umdj.edu/nigms>). Each cell strain has been hybridized onto a CGH-array of 2276 BAC's spotted in triplicate.

**Usage**

```
data(snijders)
```

**Source**

[http://www.nature.com/ng/journal/v29/n3/suppinfo/ng754\\_S1.html](http://www.nature.com/ng/journal/v29/n3/suppinfo/ng754_S1.html)

**References**

A M Snijders, N Nowak, R Segreaves, S Blackwood, N Brown, J Conroy, G Hamilton, A K Hindle, B Huey, K Kimura, S Law, K Myambo, J Palmer, B Ylstra, J P Yue, J W Gray, A N Jain, D Pinkel & D G Albertson, Assembly of microarrays for genome-wide measurement of DNA copy number, *Nature Genetics* 29, pp 263 - 264 (2001) Brief Communications.

**Examples**

```
data(snijders)
array <- gm13330
```

---

`tkdaglad`*Graphical interface for GLAD package*

---

**Description**

A graphical interface to analyse array CGH data.

**Arguments**

`list`            A character vector with the array to be analysed

**Note**

People interested in tools dealing with array CGH analysis can visit our web-page <http://bioinfo.curie.fr>.

**Author(s)**

Philippe Hupé, <glad@curie.fr>.

**See Also**

[glad](#), [daglad](#), [plotProfile](#).

**Examples**

```
data(snijders)
array1 <- as.profileCGH(gm13330)
array2 <- as.profileCGH(gm04435)

## tkdaglad(c("array1", "array2"))
## tkglad(c("array1", "array2"))
```

---

`veltman`*Public CGH data of Veltman*

---

**Description**

The data consist of 2 bladder cancer tumors obtained by Veltman et al (2003).

**Usage**

```
data(veltman)
```

**Source**

<http://cancerres.aacrjournals.org/cgi/content/full/63/11/2872>

**References**

Joris A. Veltman, Jane Fridlyand, Sunanda Pejavar, Adam B. Olshen, James E. Korkola, Sandy DeVries, Peter Carroll, Wen-Lin Kuo, Daniel Pinkel, Donna Albertson, Carlos Cordon-Cardo, Ajay N. Jain and Frederic M. Waldman. Array-based Comparative Genomic Hybridization for Genome-Wide Screening of DNA Copy Number in Bladder Tumors. *Cancer Research* 63, 2872-2880, 2003.

**Examples**

```
data(veltman)  
P9
```

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