

Package ‘flowBeads’

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Type Package

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Title flowBeads: Analysis of flow bead data

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Description This package extends flowCore to provide functionality specific to bead data. One of the goals of this package is to automate analysis of bead data for the purpose of normalisation.

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Depends R (>= 2.15.0), methods, Biobase, rrcov, flowCore

Imports flowCore, rrcov, knitr, xtable

Suggests flowViz

License Artistic-2.0

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'show-methods.R' 'plot-methods.R' 'beads.R' 'beads1-data.R'
'beads2-data.R' 'dakomef-data.R' 'cytoalmef-data.R'
'flowBeads-package.R'

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R topics documented:

flowBeads-package	2
absoluteNormalise	3
BeadFlowFrame-class	3
beads1	4
beads2	4
cytoalmef	4
dakomef	4
gateBeads	5
GatedBeadFlowFrame-class	5
generateReport	6
getClusteringStats	6
getDate	6
getMEFparams	6
getMEFtransform	7
getParams	7
getTransformFunction	7
hasMEF	7
length	8
mefTransform	8
plot	8
relativeNormalise	9
show	9
toMEF	9
Index	10

flowBeads-package *flowBeads*

Description

Bioconductor package for working with calibration beads in flow cytometry. Based on flowCore package.

Author(s)

Nikolas Pontikos <n.pontikos@gmail.com>

absoluteNormalise	<i>absoluteNormalise</i>
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Description

Absolute normalise to align peaks of bead.data to MEF.

Arguments

bead.data	GatedBeadFlowFrame
mef.data	data.frame

Value

A list of affine functions from transformed MFI relative coordinates to transformed MEF absolute coordinates.

BeadFlowFrame-class	<i>BeadFlowFrame</i>
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Description

Extension of [flowFrame](#) specific for bead data.

The constructor take as arguments the FCS file and the file containing the MEF values of the beads on the different detector channels

Usage

```
BeadFlowFrame(fcs.filename, bead.filename)
```

Arguments

fcs.filename	The file name of the FCS to load. File is loaded with the read.FCS function.
bead.filename	The file name of the MEF configuration files indicating the type of beads in the FCS file. The bead.file is read with read.csv .

Slots

fcs.filename: The file name of the FCS file from which to read.

bead.filename: The file name of the bead config file.

beads.mef: The [data.frame](#) containing the MEF of the bead populations on different channels.

trans: The transform f to linearise the fluorescence.

inv.trans: The inverse transform of f^{-1} .

beads1	<i>Dako beads on day 1</i>
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Description

Dako beads on day 1

beads2	<i>Dako beads on day 2</i>
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Description

Dako beads on day 2

cytocalmef	<i>Cytocal config file</i>
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Description

Cytocal config file

dakomef	<i>Dako config file</i>
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Description

Dako config file

`gateBeads`*gateBeads*

Description

`gateBeads` gates on all channels, apply scatter gate first. Find parameters in MEF data.frame which are also present in `BeadFlowFrame` The number of expected bead populations is by default six and it is assumed that there is the same number of beads in each population.

Arguments

<code>bead.data</code>	The <code>BeadFlowFrame</code> object to gate.
<code>K</code>	The number of bead populations expected.
<code>verbose</code>	Whether to print debug information.

Value

[GatedBeadFlowFrame](#)

Examples

```
data(beads1)
gateBeads(beads1)
```

`GatedBeadFlowFrame-class`*GatedBeadFlowFrame*

Description

`GatedBeadFlowFrame`

Arguments

<code>labels</code>	The resulting labels of the clustering assigning each event to a different bead population.
<code>clustering.stats</code>	Three dimensional array summarising the stats per channel and population.
<code>mef.transform</code>	The list of MEF transforms

generateReport *generateReport*

Description

Generate an HTML report from a Markdown template using [knitr](#).

Arguments

bead.data [GatedBeadFlowFrame](#)
output.file name of the file to which to output the HTML report.

See Also

[knitr](#)

getClusteringStats *getClusteringStats*

Description

Returns clustering stats as a 3-dimensional array.

getDate *getDate*

Description

getDate

Arguments

flow.frame [flowFrame](#) object on which to get the date field

getMEFparams *getMEFparams*

Description

Returns all the MEF parameter names.

`getMEFtransform` *getMEFtransform*

Description

Returns MEF transform function.

`getParams` *getParams*

Description

Returns all the parameter names except the scatter channels.

`getTransformFunction` *getTransformFunction*

Description

Returns transform function. The default is the logicle transform for FCS 3 and the log10 transform for FCS 2.

`hasMEF` *hasMEF*

Description

Checks whether we have the MEF for a channel name.

Arguments

`bead.data` [BeadFlowFrame](#)
`parameter` [character](#)

length	<i>length</i>
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Description

Returns the number of events in a `flowFrame` object.

Arguments

`flow.frame` `flowFrame` object on which to get number of beads

mefTransform	<i>Logicle transformation constructor</i>
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Description

Input parameters are to be provided in decades

Usage

```
mefTransform(transformationId = "mefTransform", alpha,
             beta)
```

Arguments

transformationId	The name of the transformation.
alpha	The intercept of the MEF transform.
beta	The slope of the MEF transform.

plot	<i>Plot the results of the clustering. Plot only the requested channel which should have a corresponding entry in the MEF files</i>
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Description

Plot the results of the clustering. Plot only the requested channel which should have a corresponding entry in the MEF files

Ungated bead data, simply draw all channels individually (no colours).

If no argument specified then plot all parameters

relativeNormalise	<i>relativeNormalise</i>
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Description

Relative normalise to align peaks of bead.data1 to those of bead.data2 Returns a list of affine functions from transformed MFI day one coordinates to transformed MFI day two coordinates. This permits comparison of channels across two days, provided the detector is stable, even in the absence of absolute MEF values.

Arguments

bead.data1: [GatedBeadFlowFrame](#) object with MFIs from day one
 bead.data2: [GatedBeadFlowFrame](#) object with MFIs from day two

Value

A list of affine functions from MFI day one coordinates to MFI day two coordinates.

show	<i>BeadFlowFrame</i>
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Description

BeadFlowFrame
 GatedBeadFlowFrame

toMEF	<i>toMEF</i>
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Description

Given bead.data and a flow.data apply the MEF transform to matching channels in flow.data.

Arguments

bead.data The GatedBeadFlowFrame object containing the MEF transform.
 flow.data The flowFrame object on which to apply the transform.

Index

- * **datasets**
 - beads1, 4
 - beads2, 4
 - cytocalmef, 4
 - dakomef, 4
- * **package**
 - flowBeads-package, 2
- absoluteNormalise, 3
- absoluteNormalise, GatedBeadFlowFrame, data.frame-method (absoluteNormalise), 3
- BeadFlowFrame, 7
- BeadFlowFrame (BeadFlowFrame-class), 3
- BeadFlowFrame-class, 3
- beads1, 4
- beads2, 4
- character, 7
- cytocalmef, 4
- dakomef, 4
- data.frame, 3
- dBeadFlowFrame (BeadFlowFrame-class), 3
- flowBeads (flowBeads-package), 2
- flowBeads-package, 2
- flowFrame, 3, 6, 8
- gateBeads, 5
- gateBeads, BeadFlowFrame-method (gateBeads), 5
- GatedBeadFlowFrame, 3, 5, 6, 9
- GatedBeadFlowFrame (GatedBeadFlowFrame-class), 5
- GatedBeadFlowFrame-class, 5
- generateReport, 6
- generateReport, GatedBeadFlowFrame, character-method (generateReport), 6
- getClusteringStats, 6
- getClusteringStats, GatedBeadFlowFrame-method (getClusteringStats), 6
- getDate, 6
- getDate, flowFrame-method (getDate), 6
- getMEFparams, 6
- getMEFparams, BeadFlowFrame-method (getMEFparams), 6
- getMEFtransform, 7
- getMEFtransform, GatedBeadFlowFrame-method (getMEFtransform), 7
- getParams, 7
- getParams, flowFrame-method (getParams), 7
- getTransformFunction, 7
- getTransformFunction, BeadFlowFrame-method (getTransformFunction), 7
- hasMEF, 7
- hasMEF, BeadFlowFrame, character-method (hasMEF), 7
- knitr, 6
- length, 8
- length, flowFrame-method (length), 8
- mefTransform, 8
- plot, 8
- plot, BeadFlowFrame, character-method (plot), 8
- plot, BeadFlowFrame, missing-method (plot), 8
- plot, GatedBeadFlowFrame, character-method (plot), 8
- read.csv, 3
- read.FCS, 3
- relativeNormalise, 9
- relativeNormalise, GatedBeadFlowFrame, GatedBeadFlowFrame-method (relativeNormalise), 9

show, [9](#)

toMEF, [9](#)

toMEF, GatedBeadFlowFrame, flowFrame-method
 (toMEF), [9](#)