

# Package ‘synapsis’

January 23, 2023

**Type** Package

**Title** An R package to automate the analysis of double-strand break repair during meiosis

**Version** 1.5.0

**Description** Synapsis is a Bioconductor software package for automated (unbiased and reproducible) analysis of meiotic immunofluorescence datasets. The primary functions of the software can i) identify cells in meiotic prophase that are labelled by a synaptonemal complex axis or central element protein, ii) isolate individual synaptonemal complexes and measure their physical length, iii) quantify foci and co-localise them with synaptonemal complexes, iv) measure interference between synaptonemal complex-associated foci. The software has applications that extend to multiple species and to the analysis of other proteins that label meiotic prophase chromosomes. The software converts meiotic immunofluorescence images into R data frames that are compatible with machine learning methods. Given a set of microscopy images of meiotic spread slides, synapsis crops images around individual single cells, counts colocalising foci on strands on a per cell basis, and measures the distance between foci on any given strand.

**biocViews** Software, SingleCell

**Depends** R (>= 4.1)

**Imports** EBImage, stats, utils, graphics

**License** MIT + file LICENSE

**Encoding** UTF-8

**RoxygenNote** 7.1.1

**VignetteBuilder** knitr

**Suggests** knitr, rmarkdown, testthat (>= 3.0.0), ggplot2, tidyverse, BiocStyle

**Config/testthat/edition** 3

**git\_url** <https://git.bioconductor.org/packages/synapsis>

**git\_branch** master

**git\_last\_commit** c110378

**git\_last\_commit\_date** 2022-11-01

**Date/Publication** 2023-01-22

**Author** Lucy McNeill [aut, cre, cph] (<<https://orcid.org/0000-0003-1752-4882>>),  
Wayne Crismani [rev, ctb] (<<https://orcid.org/0000-0003-0143-8293>>)

**Maintainer** Lucy McNeill <luc.mcneill@gmail.com>

## R topics documented:

annotate_foci_counting . . . . .	2
annotate_foci_counting_adjusted . . . . .	3
append_data_frame . . . . .	4
auto_crop_fast . . . . .	5
count_foci . . . . .	8
crop_single_object_fast . . . . .	10
get_blobs . . . . .	12
get_C1 . . . . .	13
get_coincident_foci . . . . .	14
get_foci_per_cell . . . . .	16
get_overlap_mask . . . . .	17
get_pachytene . . . . .	18
keep_cells . . . . .	19
make_foci_mask . . . . .	20
make_strand_mask . . . . .	21
remove_XY . . . . .	22

<b>Index</b>	<b>23</b>
--------------	-----------

---

annotate\_foci\_counting  
*annotate\_foci\_counting*

---

### Description

Contains all plotting routines for count foci annotation

### Usage

```

annotate_foci_counting(
  img_file,
  cell_count,
  img_orig,
  img_orig_foci,
  artificial_amp_factor,
  strands,
  coincident_foci,
  foci_label,
  alone_foci,
  percent_px,
  foci_per_cell
)

```

**Arguments**

img_file	cell's file name
cell_count	unique cell counter
img_orig	original strand crop
img_orig_foci	cropped foci channel
artificial_amp_factor	amplification factor
strands	black white mask of strand channel
coincident_foci	mask of overlap between strand and foci channel
foci_label	black and white mask of foci channel
alone_foci	estimated number of foci that are NOT on a strand.
percent_px	percentage of foci mask that coincides with strand channel small number indicates potentially problematic image.
foci_per_cell	number of foci counted per cell

**Value**

displays key steps from raw image to coincident foci count

---

annotate\_foci\_counting\_adjusted  
*annotate\_foci\_counting\_adjusted*

---

**Description**

Contains all plotting routines for count foci annotation

**Usage**

```

annotate_foci_counting_adjusted(
  img_file,
  cell_count,
  img_orig,
  img_orig_foci,
  artificial_amp_factor,
  strands,
  coincident_foci,
  foci_label,
  alone_foci,
  percent_px,
  foci_per_cell
)

```

**Arguments**

img_file	cell's file name
cell_count	unique cell counter
img_orig	original strand crop
img_orig_foci	cropped foci channel
artificial_amp_factor	amplification factor
strands	black white mask of strand channel
coincident_foci	mask of overlap between strand and foci channel
foci_label	black and white mask of foci channel
alone_foci	estimated number of foci that are NOT on a strand.
percent_px	percentage of foci mask that coincides with strand channel small number indicates potentially problematic image.
foci_per_cell	number of foci counted per cell

**Value**

displays key steps from raw image to coincident foci count

---

append\_data\_frame      *append\_data\_frame*

---

**Description**

applies new row to data frame

**Usage**

```
append_data_frame(
  WT_str,
  KO_str,
  WT_out,
  KO_out,
  img_file,
  foci_areas,
  df_cells,
  cell_count,
  stage,
  foci_per_cell,
  image_mat,
  percent_px,
  alone_foci,
  discrepant_category,
  C1
)
```

**Arguments**

WT_str	string in filename corresponding to wildtype genotype. Defaults to ++.
KO_str	string in filename corresponding to knockout genotype. Defaults to -.
WT_out	string in output csv in genotype column, for knockout. Defaults to +/+.
KO_out	string in output csv in genotype column, for knockout. Defaults to -/-.
img_file	cell's file name
foci_areas	pixel area of each foci
df_cells	current data frame
cell_count	unique cell counter
stage,	meiosis stage of interest. Currently count_foci determines this with thresholding/ object properties in the synaptonemal complex channel by previously calling the get_pachytene function. Note that if using this option, the count_foci function requires that the input directory contains a folder called "pachytene" with the crops in it.
foci_per_cell	foci count for cell
image_mat	matrix with all pixel values above zero
percent_px	percentage of foci mask that coincides with strand channel small number indicates potentially problematic image.
alone_foci	estimated number of foci that are NOT on a strand.
discrepant_category	estimated number of foci that are NOT on a strand.
C1	criteria

**Value**

data frame with new row

---

auto_crop_fast	<i>auto_crop_fast</i>
----------------	-----------------------

---

**Description**

crop an image around each viable cell candidate.

**Usage**

```
auto_crop_fast(
  img_path,
  max_cell_area = 20000,
  min_cell_area = 7000,
  mean_pix = 0.08,
  annotation = "off",
```

```

blob_factor = 15,
bg_blob_factor = 10,
offset = 0.2,
final_blob_amp = 10,
test_amount = 0,
brush_size_blob = 51,
sigma_blob = 15,
channel3_string = "DAPI",
channel2_string = "SYCP3",
channel1_string = "MLH3",
file_ext = "jpeg",
third_channel = "off",
cell_aspect_ratio = 2,
strand_amp = 2,
path_out = img_path,
resize_l = 720,
crowded_cells = "FALSE",
watershed_radius = 50,
watershed_tol = 0.2,
cropping_factor = 1.3
)

```

### Arguments

img_path,	path containing image data to analyse
max_cell_area,	Maximum pixel area of a cell candidate
min_cell_area,	Minimum pixel area of a cell candidate
mean_pix,	Mean pixel intensity of cell crop (in SYCP3 channel) for normalisation
annotation,	Choice to output pipeline choices (recommended to knit)
blob_factor,	Contrast factor to multiply original image by before smoothing/smudging
bg_blob_factor,	Contrast factor to multiply original image by to take background. Used prior to thresholding.
offset,	Pixel value offset from bg_blob_factor. Used in thresholding to make blob mask.
final_blob_amp,	Contrast factor to multiply smoothed/smudged image. Used in thresholding to make blob mask.
test_amount,	Optional number of first N images you want to run function on. For troubleshooting/testing/variable calibration purposes.
brush_size_blob,	Brush size for smudging the synaptonemal complex channel to make blobs
sigma_blob,	Sigma in Gaussian brush for smudging the synaptonemal complex channel to make blobs

channel3_string	Optional. String appended to the files showing the channel illuminating cell structures. Defaults to DAPI, if third channel == "on".
channel2_string	String appended to the files showing the channel illuminating synaptonemal complexes. Defaults to SYCP3
channel1_string	String appended to the files showing the channel illuminating foci. Defaults to MLH3
file_ext	file extension of your images e.g. tif jpeg or png.
third_channel	Optional, defaults to "off". Set to "on" if you would also like crops of the third channel.
cell_aspect_ratio	Maximum aspect ratio of blob to be defined as a cell
strand_amp	multiplication of strand channel for get_blobs function.
path_out,	user specified output path. Defaults to img_path
resize_l	length for resized image
crowded_cells	TRUE or FALSE, defaults to FALSE. Set to TRUE if you have many cells in a frame that almost touch
watershed_radius	Radius (ext variable) in watershed method used in strand channel. Defaults to 1 (small)
watershed_tol	Intensity tolerance for watershed method. Defaults to 0.05.
cropping_factor	size of cropping window square, as factor of characteristic blob radius. Defaults to 1. May need to increase if using watershed.

## Details

This function takes all images in a directory, and crops around individual cells according to the antibody that stains synaptonemal complexes e.g. SYCP3. First, it increases the brightness and smudges the image with a Gaussian brush, and creates a mask using thresholding (get\_blobs). Then it deletes cell candidates in the mask deemed too large, too small, or too long (keep\_cells). Using the computeFeatures functions from EBImage to locate centre and radius, the cropping area is determined and the original image cropped. These images are saved in either a user specified directory, or a crops folder at the location of the image files.

## Value

cropped synaptonemal complex and foci channels around single cells, regardless of stage

## Author(s)

Lucy McNeill

**Examples**

```
demo_path = paste0(system.file("extdata", package = "synapsis"))
auto_crop_fast(demo_path, annotation = "on", max_cell_area = 30000,
min_cell_area = 7000, file_ext = "tif", crowded_cells = TRUE)
```

---

count\_foci

*count\_foci*


---

**Description**

Calculates coincident foci in synaptonemal complex and foci channel, per cell

**Usage**

```
count_foci(
  img_path,
  stage = "none",
  offset_px = 0.2,
  offset_factor = 2,
  brush_size = 3,
  brush_sigma = 3,
  foci_norm = 0.01,
  annotation = "off",
  channel2_string = "SYCP3",
  channel1_string = "MLH3",
  file_ext = "jpeg",
  KO_str = "--",
  WT_str = "++",
  KO_out = "-/-",
  WT_out = "+/+ ",
  watershed_stop = "off",
  watershed_radius = 1,
  watershed_tol = 0.05,
  crowded_foci = TRUE,
  artificial_amp_factor = 1,
  strand_amp = 2,
  min_foci = -1,
  disc_size = 51,
  modify_problematic = "off",
  disc_size_foci = 5,
  C1 = 0.02,
  C2 = 0.46,
  C_weigh_foci_number = TRUE
)
```



**Arguments**

img_path,	path containing crops to analyse
stage,	meiosis stage of interest. Currently count_foci determines this with thresholding/ object properties in the synaptonemal complex channel by previously calling the get_pachytene function. Note that if using this option, the count_foci function requires that the input directory contains a folder called "pachytene" with the crops in it.
offset_px,	Pixel value offset used in thresholding of synaptonemal complex channel
offset_factor,	Pixel value offset used in thresholding of foci channel
brush_size,	size of brush to smooth the foci channel. Should be small to avoid erasing foci.
brush_sigma,	sigma for Gaussian smooth of foci channel. Should be small to avoid erasing foci.
foci_norm,	Mean intensity to normalise all foci channels to.
annotation,	Choice to output pipeline choices (recommended to knit)
channel2_string	String appended to the files showing the channel illuminating synaptonemal complexes. Defaults to SYCP3
channel1_string	String appended to the files showing the channel illuminating foci. Defaults to MLH3
file_ext	file extension of your images e.g. tiff jpeg or png.
KO_str	string in filename corresponding to knockout genotype. Defaults to -.
WT_str	string in filename corresponding to wildtype genotype. Defaults to ++.
KO_out	string in output csv in genotype column, for knockout. Defaults to -/-.
WT_out	string in output csv in genotype column, for knockout. Defaults to +/-.
watershed_stop	Stop default watershed method with "on"
watershed_radius	Radius (ext variable) in watershed method used in foci channel. Defaults to 1 (small)
watershed_tol	Intensity tolerance for watershed method. Defaults to 0.05.
crowded_foci	TRUE or FALSE, defaults to FALSE. Set to TRUE if you have foci > 100 or so.
artificial_amp_factor	Amplification of foci channel, for annotation only.
strand_amp	multiplication of strand channel to make masks
min_foci	minimum pixel area for a foci. Depends on your dpi etc. Defaults to 4
disc_size	size of disc for local background calculation in synaptonemal complex channel
modify_problematic	option for synapsis to try and "save" images which have likely been counted incorrectly due to a number of reasons. Default settings are optimized for mouse pachytene. Defaults to "off"

`disc_size_foci` size of disc for local background calculation in foci channel  
`C1` Default crispness criteria =  $\text{sd}(\text{foci\_area})/(\text{mean}(\text{foci\_area})+1)$   
`C2` Alternative crisp criteria.  
`C_weigh_foci_number` choose crispness criteria- defaults to TRUE to use C1 (weighing with number).  
 Otherwise set to FALSE to use C2

### Details

In this function, masks for the synaptonemal complex (SC) and foci channel are created from the saved crops of single/individual cells. These masks are computed using (optional) input parameters related to meiosis stage/ how well spread chromosomes are (for the former) and related to smoothing, thresholding and how "crowded" foci are for the latter. Finally, these two masks are multiplied, and the number of objects found with EImage's `computeFeatures` are the colocalizing foci.

The file, cell number, foci count etc. are output as a data frame.

### Value

data frame with foci count per cell

### Author(s)

Lucy McNeill

### Examples

```

demo_path = paste0(system.file("extdata", package = "synapsis"))
foci_counts <- count_foci(demo_path, offset_factor = 3, brush_size = 3,
brush_sigma = 3, annotation = "on", stage = "pachytene")
  
```

---

`crop_single_object_fast`  
*crop\_single\_object\_fast*

---

### Description

Creates mask for every individual cell candidate in mask

### Usage

```

crop_single_object_fast(
  retained,
  OOI_final,
  counter_final,
  img_orig,
  img_orig_foci,
  img_orig_DAPI = "blank",
  )
  
```

```

    file_sc,
    file_foci,
    file_DAPI = "blank",
    cell_count,
    mean_pix,
    annotation,
    file_base,
    img_path,
    r_max,
    cx,
    cy,
    channel3_string,
    channel2_string,
    channel1_string,
    file_ext,
    third_channel,
    path_out,
    img_orig_highres,
    resize_l,
    crowded_cells,
    cropping_factor
)

```

### Arguments

retained	Mask of cell candidates which meet size criteria. After smoothing/smudging and thresholding.
OOI_final,	Objects of interest count. Total number of cell candidates in retained.
counter_final,	Counter for single cell we are focussing on. Remove all other cells where counter_single not equal to counter_final.
img_orig,	description
img_orig_foci,	description
img_orig_DAPI,	description
file_sc,	filename of synaptonemal complex channel image
file_foci,	filename of foci channel image
file_DAPI,	filename of DAPI channel image
cell_count,	counter for successful crops around cells
mean_pix,	Mean pixel intensity of cell crop (in SYCP3 channel) for normalisation
annotation,	Choice to output pipeline choices (recommended to knit)
file_base,	filename base common to all three channels i.e. without -MLH3.jpeg etc.
img_path,	path containing image data to analyse
r_max	maximum radius of blob for cropping

<code>cx</code>	centre of blob x
<code>cy</code>	centre of blob y
<code>channel3_string</code>	Optional. String appended to the files showing the channel illuminating cell structures. Defaults to DAPI, if third channel == "on".
<code>channel2_string</code>	String appended to the files showing the channel illuminating synaptonemal complexes. Defaults to SYCP3
<code>channel1_string</code>	String appended to the files showing the channel illuminating foci. Defaults to MLH3
<code>file_ext</code>	file extension of your images e.g. tif jpeg or png.
<code>third_channel</code>	Optional, defaults to "off". Set to "on" if you would also like crops of the third channel.
<code>path_out,</code> <code>img_orig_highres</code>	user specified output path. Defaults to <code>img_path</code> the original strand image with original resolution
<code>resize_l</code>	length of square to resize original image to.
<code>crowded_cells</code>	TRUE or FALSE, defaults to FALSE. Set to TRUE if you have many cells in a frame that almost touch
<code>cropping_factor</code>	size of cropping window square, as factor of characteristic blob radius. Defaults to 1. May need to increase if using watershed.

**Value**

Crops around all candidates in both channels

---

`get_blobs`

*get\_blobs*

---

**Description**

Makes mask of all objects bright enough to be classified as a cell candidate

**Usage**

```
get_blobs(
  img_orig,
  blob_factor,
  bg_blob_factor,
  offset,
  final_blob_amp,
  brush_size_blob,
```

```

    sigma_blob,
    watershed_tol,
    watershed_radius,
    crowded_cells,
    annotation
)

```

### Arguments

img_orig	Original image
blob_factor,	Contrast factor to multiply original image by before smoothing/smudging
bg_blob_factor,	Contrast factor to multiply original image by to take background. Used prior to thresholding.
offset,	Pixel value offset from bg_blob_factor. Used in thresholding to make blob mask.
final_blob_amp,	Contrast factor to multiply smoothed/smudged image. Used in thresholding to make blob mask.
brush_size_blob,	Brush size for smudging the synaptonemal complex channel to make blobs
sigma_blob,	Sigma in Gaussian brush for smudging the synaptonemal complex channel to make blobs
watershed_tol	Intensity tolerance for watershed method. Defaults to 0.05.
watershed_radius	Radius (ext variable) in watershed method used in strand channel. Defaults to 1 (small)
crowded_cells	TRUE or FALSE, defaults to FALSE. Set to TRUE if you have many cells in a frame that almost touch
annotation,	Choice to output pipeline choices (recommended to knit) have many cells in a frame that almost touch

### Value

Mask with cell candidates

---

get_C1	<i>get_C1</i>
--------	---------------

---

### Description

calculates the statistic to compare to `crisp_criteria`, which determines whether the foci count will be reliable

### Usage

```
get_C1(foci_areas, foci_per_cell, C_weigh_foci_number)
```

**Arguments**

foci\_areas      pixel area of each foci  
foci\_per\_cell   foci count for cell  
C\_weigh\_foci\_number  
                 choose crispness criteria- defaults to TRUE to use C1 (weighing with number).  
                 Otherwise set to FALSE to use C2

**Value**

statistic to compare to `crisp_criteria`

---

`get_coincident_foci`    *get\_coincident\_foci*

---

**Description**

calculates the statistic to compare to `crisp_criteria`, which determines whether the foci count will be reliable

**Usage**

```
get_coincident_foci(  
  offset_px,  
  offset_factor,  
  brush_size,  
  brush_sigma,  
  annotation,  
  watershed_stop,  
  watershed_radius,  
  watershed_tol,  
  crowded_foci,  
  artificial_amp_factor,  
  strand_amp,  
  disc_size,  
  disc_size_foci,  
  img_file,  
  cell_count,  
  img_orig,  
  img_orig_foci,  
  stage,  
  WT_str,  
  KO_str,  
  WT_out,  
  KO_out,  
  C1_search,  
  discrepant_category,
```

```

    C1,
    C2,
    df_cells,
    C_weigh_foci_number
)

```

### Arguments

offset_px,	Pixel value offset used in thresholding of synaptonemal complex channel
offset_factor,	Pixel value offset used in thresholding of foci channel
brush_size,	size of brush to smooth the foci channel. Should be small to avoid erasing foci.
brush_sigma,	sigma for Gaussian smooth of foci channel. Should be small to avoid erasing foci.
annotation,	Choice to output pipeline choices (recommended to knit)
watershed_stop	Stop default watershed method with "on"
watershed_radius	Radius (ext variable) in watershed method used in foci channel. Defaults to 1 (small)
watershed_tol	Intensity tolerance for watershed method. Defaults to 0.05.
crowded_foci	TRUE or FALSE, defaults to FALSE. Set to TRUE if you have foci > 100 or so.
artificial_amp_factor	Amplification of foci channel, for annotation only.
strand_amp	multiplication of strand channel to make masks
disc_size	size of disc for local background calculation in synaptonemal complex channel
disc_size_foci	size of disc for local background calculation in foci channel
img_file	cell's file name
cell_count	unique cell counter
img_orig	original strand crop
img_orig_foci	cropped foci channel
stage,	meiosis stage of interest. Currently count_foci determines this with thresholding/ object properties in the synaptonemal complex channel by previously calling the get_pachytene function. Note that if using this option, the count_foci function requires that the input directory contains a folder called "pachytene" with the crops in it.
WT_str	string in filename corresponding to wildtype genotype. Defaults to ++.
KO_str	string in filename corresponding to knockout genotype. Defaults to --.
WT_out	string in output csv in genotype column, for knockout. Defaults to +/+.
KO_out	string in output csv in genotype column, for knockout. Defaults to -/-.
C1_search	TRUE or FALSE whether the image is still being modified until it meets the crispness criteria

discrepant\_category      estimated number of foci that are NOT on a strand.  
 C1                          Default crispness criteria =  $sd(foci\_area)/(mean(foci\_area)+1)$   
 C2                          Alternative crisp criteria.  
 df\_cells                  current data frame  
 C\_weigh\_foci\_number      choose crispness criteria- defaults to TRUE to use C1 (weighing with number).  
                                  Otherwise set to FALSE to use C2

**Value**

data frame with new row with most recent foci per cell appended

---

*get\_foci\_per\_cell*      *get\_foci\_per\_cell*

---

**Description**

creates mask for coincident foci

**Usage**

```

get_foci_per_cell(
  img_file,
  offset_px,
  stage,
  strands,
  watershed_stop,
  foci_label,
  annotation,
  cell_count,
  img_orig,
  img_orig_foci,
  artificial_amp_factor,
  coincident_foci
)

```

**Arguments**

img\_file                  cell's file name  
 offset\_px,                Pixel value offset used in thresholding of synaptonemal complex channel  
 stage,                    meiosis stage of interest. Currently count\_foci determines this with thresholding/ object properties in the synaptonemal complex channel by previously calling the get\_pachytene function. Note that if using this option, the count\_foci function requires that the input directory contains a folder called "pachytene" with the crops in it.



strands           black white mask of strand channel  
 watershed\_stop   Stop default watershed method with "on"  
 foci\_label        black and white mask of foci channel  
 annotation,       Choice to output pipeline choices (recommended to knit)  
 cell\_count        unique cell counter  
 img\_orig          original strand crop  
 img\_orig\_foci    cropped foci channel  
 artificial\_amp\_factor  
                   amplification factor  
 coincident\_foci  
                   mask of coincident foci

**Value**

number of foci per cell

---

`get_overlap_mask`        *get\_overlap\_mask*

---

**Description**

creates mask for coincident foci

**Usage**

```

get_overlap_mask(
  strands,
  foci_label,
  watershed_stop,
  img_orig_foci,
  watershed_radius,
  watershed_tol
)

```

**Arguments**

strands           black white mask of strand channel  
 foci\_label        black and white mask of foci channel  
 watershed\_stop   Stop default watershed method with "on"  
 img\_orig\_foci    cropped foci channel  
 watershed\_radius  
                   Radius (ext variable) in watershed method used in foci channel. Defaults to 1  
                   (small)  
 watershed\_tol    Intensity tolerance for watershed method. Defaults to 0.05.

**Value**

mask with coincident foci on strands

---

<code>get_pachytene</code>	<i>get_pachytene</i>
----------------------------	----------------------

---

**Description**

Identifies crops in pachytene

**Usage**

```
get_pachytene(
  img_path,
  species_num = 20,
  offset = 0.2,
  ecc_thresh = 0.85,
  area_thresh = 0.06,
  annotation = "off",
  channel2_string = "SYCP3",
  channel1_string = "MLH3",
  file_ext = "jpeg",
  KO_str = "--",
  WT_str = "++",
  KO_out = "-/-",
  WT_out = "+/+ ",
  path_out = img_path,
  artificial_amp_factor = 3,
  strand_amp = 2,
  resize_l = 120
)
```

**Arguments**

<code>img_path</code> ,	path containing crops analyse
<code>species_num</code> ,	number of chromosomes in the species
<code>offset</code> ,	Pixel value offset used in thresholding for the synaptonemal complex (SYCP3) channel
<code>ecc_thresh</code> ,	The minimum average eccentricity of all objects in mask determined by computefeatures, for a cell to be pachytene.
<code>area_thresh</code> ,	The minimum ratio of pixels included in mask to total, for a cell to be classified as pachytene.
<code>annotation</code> ,	Choice to output pipeline choices (recommended to knit)
<code>channel2_string</code>	String appended to the files showing the channel illuminating synaptonemal complexes. Defaults to SYCP3

channel_string	String appended to the files showing the channel illuminating foci. Defaults to MLH3
file_ext	file extension of your images e.g. tiff jpeg or png.
KO_str	string in filename corresponding to knockout genotype. Defaults to -.
WT_str	string in filename corresponding to wildtype genotype. Defaults to ++.
KO_out	string in output csv in genotype column, for knockout. Defaults to -/-.
WT_out	string in output csv in genotype column, for knockout. Defaults to +/-.
path_out,	user specified output path. Defaults to img_path
artificial_amp_factor	Amplification of foci channel, for RGB output files. Defaults to 3.
strand_amp	multiplication of strand channel.
resize_l	length of resized square cell image.

### Details

This function takes the crops made by `auto_crop` fast, and determines the number of synaptonemal complex candidates by considering the local background and using EBImage functions. In general, very bright objects which contrast highly with the background will be classified as the same object. Dim objects will likely be classified as many different objects. If the number of objects is too high compared to the species number (`species_num`) then the cell is determined to not be in pachytene. Note that this function has been optimized for mouse cells which can be very well spread / separated.

### Value

Pairs of foci and synaptonemal channel crops for pachytene

### Author(s)

Lucy McNeill

### Examples

```
demo_path = paste0(system.file("extdata", package = "synapsis"))
SYCP3_stats <- get_pachytene(demo_path, ecc_thresh = 0.8, area_thresh = 0.04, annotation = "on")
```

---

keep_cells	<i>keep_cells</i>
------------	-------------------

---

### Description

Deletes objects in mask which are too small, large, oblong i.e. unlikely to be a cell

**Usage**

```
keep_cells(
  candidate,
  max_cell_area,
  min_cell_area,
  cell_aspect_ratio,
  crowded_cells,
  annotation
)
```

**Arguments**

candidate	Mask of individual cell candidates
max_cell_area,	Maximum pixel area of a cell candidate
min_cell_area,	Minimum pixel area of a cell candidate
cell_aspect_ratio	Maximum aspect ratio of blob to be defined as a cell
crowded_cells	TRUE or FALSE, defaults to FALSE. Set to TRUE if you
annotation,	Choice to output pipeline choices (recommended to knit) have many cells in a frame that almost touch

**Value**

Mask of cell candidates which meet size criteria

---

make_foci_mask	<i>make_foci_mask</i>
----------------	-----------------------

---

**Description**

creates foci mask for foci channel crop

**Usage**

```
make_foci_mask(
  offset_factor,
  bg,
  crowded_foci,
  img_orig_foci,
  brush_size,
  brush_sigma,
  disc_size_foci
)
```

**Arguments**

offset_factor	Pixel value offset used in thresholding of foci channel
bg	background value- currently just mean pixel value of whole image
crowded_foci	TRUE or FALSE, defaults to FALSE. Set to TRUE if you have foci > 100 or so.
img_orig_foci	cropped foci channel
brush_size	size of brush to smooth the foci channel. Should be small to avoid erasing foci.
brush_sigma	sigma for Gaussian smooth of foci channel. Should be small to avoid erasing foci.
disc_size_foci	size of disc for local background calculation in foci channel

**Value**

foci mask

---

make_strand_mask	<i>make_strand_mask</i>
------------------	-------------------------

---

**Description**

creates strand mask for strand channel crop

**Usage**

```
make_strand_mask(
  offset_px,
  stage,
  img_orig,
  disc_size,
  brush_size,
  brush_sigma
)
```

**Arguments**

offset_px,	Pixel value offset used in thresholding of synaptonemal complex channel
stage,	meiosis stage of interest. Currently count_foci determines this with thresholding/ object properties in the synaptonemal complex channel by previously calling the get_pachytene function. Note that if using this option, the count_foci function requires that the input directory contains a folder called "pachytene" with the crops in it.
img_orig	original strand crop
disc_size	size of disc for local background calculation in synaptonemal complex channel
brush_size,	size of brush to smooth the foci channel. Should be small to avoid erasing foci.
brush_sigma,	sigma for Gaussian smooth of foci channel. Should be small to avoid erasing foci.

**Value**

strand mask

---

remove\_XY

*remove\_XY*

---

**Description**

applies new row to data frame

**Usage**

```
remove_XY(foci_label, foci_candidates, foci_areas)
```

**Arguments**

foci\_label      black and white mask of foci channel  
foci\_candidates  
                 computeFeatures data frame of foci channel  
foci\_areas      the areas of the foci objects

**Value**

mask with XY blob removed

# Index

[annotate\\_foci\\_counting](#), 2  
[annotate\\_foci\\_counting\\_adjusted](#), 3  
[append\\_data\\_frame](#), 4  
[auto\\_crop\\_fast](#), 5

[count\\_foci](#), 8  
[crop\\_single\\_object\\_fast](#), 10

[get\\_blobs](#), 12  
[get\\_C1](#), 13  
[get\\_coincident\\_foci](#), 14  
[get\\_foci\\_per\\_cell](#), 16  
[get\\_overlap\\_mask](#), 17  
[get\\_pachytene](#), 18

[keep\\_cells](#), 19

[make\\_foci\\_mask](#), 20  
[make\\_strand\\_mask](#), 21

[remove\\_XY](#), 22